

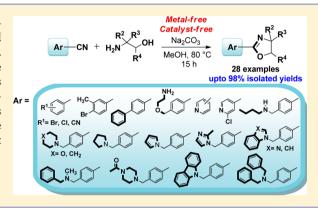
# Synthesis of 2-Aryl/Heteroaryloxazolines from Nitriles under Metaland Catalyst-Free Conditions and Evaluation of Their Antioxidant **Activities**

Parul Garg, Shweta Chaudhary, and Marilyn D. Milton\*

Department of Chemistry, University of Delhi, Delhi 110 007, India

Supporting Information

ABSTRACT: The synthesis of structurally diverse 2-aryl/heteroaryloxazolines from nitriles and aminoalcohols has been achieved under metal- and catalyst-free conditions in good to excellent yields. An array of functional groups are well-tolerated, thus, allowing the introduction of many important biologically active motifs such as azoles, ring-fused azoles, saturated heterocyclics, and amines in 2aryloxazoline scaffolds. An evaluation of the antioxidant properties using the DPPH (diphenyl picryl hydrazyl) assay method shows the pyrrole-functionalized 2-aryloxazoline to be the best antioxidant among all the synthesized 2-aryl/heteroaryloxazolines.



#### ■ INTRODUCTION

2-Aryloxazoline scaffolds have been the core structures of many biologically active molecules that exhibit cytotoxic, <sup>1a</sup> antibacterial, <sup>1b</sup> antitumor, <sup>1c,d</sup> antidepressant, <sup>1e,f</sup> anti-Alzheimer's, <sup>1g</sup> or deferrating<sup>1h</sup> activities (Figure 1). Also, these moieties are used as synthetic intermediates,<sup>2</sup> protecting groups,<sup>3</sup> ligands,<sup>4</sup> or chiral auxiliaries<sup>5</sup> in many organic transformations. Furthermore, many polyoxazolines are useful biomaterials for drug and gene delivery, and stimuli-responsive systems.<sup>6</sup> In addition, oxazolines can also be dehydrogenated to produce oxazoles, another important class of bioactive molecules. Recently, the antioxidant properties of aryl-substituted 2oxazolines have also been explored.8

A number of chemical methods have been developed for the synthesis of 2-aryl/heteroaryloxazoline scaffolds using  $\beta$ hydroxyl amides, olefins, carboxylic acids, esters, nitriles, aldehydes, or other carbonyl-containing compounds. <sup>9–12</sup> The transition-metal-catalyzed methods such as carbonylative cyclization of aryl bromides with 2-chloroethanamine 10 or direct arylation of C-H bond of nonaromatic oxazolines<sup>11</sup> often require long reaction hours at high reaction temperatures (>100 °C) and have limited substrate scope. Several catalysts such as ZnCl<sub>2</sub>, Bi (III) salts, ZrOCl<sub>2</sub>·8H<sub>2</sub>O, clay, cellulose sulfuric acid, silica sulfuric acid, H<sub>3</sub>PW<sub>12</sub>O<sub>40</sub>, Cu complexes, and S-Co (II) salts have been used for the activation of nitriles toward nucleophilic addition by aminoalcohols. 12 Although the aforementioned methods are promising synthetic strategies for 2-aryl/heteroaryloxazolines, however, lack of general applicability, use of less abundant and expensive reagents or catalysts, harsh reaction conditions, high reaction temperatures, or long reaction hours limit their application. Moreover, contamination

of drug-related precursors with even traces of transition metals can cause severe problems. As a result, there is a need to develop a general, cost-effective, and metal-free procedure for the synthesis of 2-aryl/heteroaryloxazolines for pharmaceutical purposes.

Herein, we describe metal- and catalyst-free condensation of nitriles with aminoalcohols for the synthesis of 2-aryl/ heteroaryloxazolines and an evaluation of their antioxidant properties. Recently, we reported sodium carbonate-promoted coupling of esters with aminoalcohols for the synthesis of N-(hydroxyalkyl)cinnamamides, <sup>13</sup> whereby reaction of ester (E)methyl p-cyanocinnamate (7) with 2-aminoethanol (8a) in the presence of Na<sub>2</sub>CO<sub>3</sub> as base in methanol afforded major amounts of (E)-N-(2-hydroxyethyl)-4-cyanocinnamamide (9)along with a minor amount of oxazoline (E)-N-(2-hydroxyethyl)-4-(4,5-dihydrooxazol-2-yl)cinnamamide (10) under aerial conditions (Scheme 1). Intrigued by direct oxazoline formation from nitrile and aminoalcohol without the use of any metal catalyst or high temperature, we decided to further optimize the reaction conditions to obtain better yields of oxazolines.

# RESULTS AND DISCUSSION

From our previous work (Scheme 1), 13 we inferred that the yield of oxazoline 10 increased upon increase in reaction time. Therefore, we began our work with the reaction of commercially available 4-bromobenzonitrile (11a) with 2aminoethanol (8a) for the synthesis of 2-(4-bromophenyl)-

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Figure 1. Examples of 2-aryl/heteroaryloxazoline-scaffold-based bioactive molecules.

# Scheme 1. Formation of Oxazoline Ring<sup>13</sup> Our previous work

NC OMe OMe OH 
$$\frac{7}{80}$$
 OMe  $\frac{10}{80}$  OH  $\frac{10}{80}$  OH  $\frac{10}{10}$  OH  $\frac{10}$ 

4,5-dihydrooxazole (12a) by employing Na<sub>2</sub>CO<sub>3</sub> as the base at 80 °C in MeOH under aerial conditions. The TLC monitoring of the reaction mixture at different time intervals showed the complete consumption of nitrile 11a in 15 h affording 47% yield of the desired product 12a together with the formation of 9% of 4-bromobenzamide (13) and 44% of 4-bromo-N-(2hydroxyethyl)benzamide (14) (Table 1, entry 1). However, performing the same reaction under nitrogen atmosphere in dry MeOH significantly improved the yield of desired oxazoline 12a to 89% with much reduced yields of byproducts 13 and 14 (Table 1, entry 2). 15c Among different types of bases screened for this reaction, DBU gave 83% yield of 12a (Table 1, entry 3) whereas other bases like NaOH, NaOMe, Et<sub>3</sub>N, K<sub>2</sub>CO<sub>3</sub>, and NaOAc furnished much lower yields (11-77%) of the desired product (Table 1, entries 4-8). Interestingly, base-free conditions afforded 80% yield of 12a along with traces of byproducts (Table 1, entry 9). Although 12a could be isolated as the almost exclusive product under base-free conditions with good yield, the presence of Na2CO3 furnished better yield of the desired product. So, we decided to use Na<sub>2</sub>CO<sub>3</sub> as the base in this transformation. Next, when we tried to reduce the reaction time from 15 to 10 h, the yield of 12a dropped to 64% (Table 1, entry 10). The use of other solvents, such as EtOH, THF, and toluene resulted in unsatisfactory yields of 12a

Table 1. Optimization of Reaction Conditions for the Synthesis of  $12a^a$ 

				yield (%) <sup>b</sup>		
entry	base	$temp\ (^{\circ}C)$	solvent	12a	13	14
1 <sup>c</sup>	Na <sub>2</sub> CO <sub>3</sub>	80	MeOH	47	9	44
2	$Na_2CO_3$	80	MeOH	89	9	<5
3	DBU	80	MeOH	83	13	<5
4	NaOH	80	MeOH	11	30	45
5	NaOMe	80	MeOH	50	41	<5
6	$Et_3N$	80	MeOH	58	13	11
7	$K_2CO_3$	80	MeOH	75	24	traces
8	NaOAc	80	MeOH	77	14	<10
9		80	MeOH	80	traces	traces
$10^d$	$Na_2CO_3$	80	MeOH	64	10	<5
11	$Na_2CO_3$	80	EtOH	55	12	14
12	$Na_2CO_3$	110	toluene	58	19	traces
13 <sup>e</sup>	$Na_2CO_3$	60	THF	15		
$14^f$	$Na_2CO_3$	80	MeOH	32	5	62
15 <sup>g</sup>	$Na_2CO_3$	r.t.	MeOH	71	traces	traces
16	$Na_2CO_3$	60	MeOH	66	10	19

"11a (1.0 mmol), 8a (5.0 mmol), base (1.0 mmol), dry solvent (1.0 mL), 80 °C, 15 h, under nitrogen atmosphere unless otherwise noted. b Isolated yields. Under aerial conditions in undried MeOH. Reaction run for 10 h. Recovery yield of 11a was 85%. J Using 3.0 mmol of 8a. Reaction carried out for 62 h.

(Table 1, entries 11–13). Furthermore, the amount of **8a** was also found to be crucial to obtain high conversions, thus, a decrease in the amount of **8a** from 5.0 to 3.0 mmol reduced the desired product yield to 32% (Table 1, entry 14). The reaction could also be performed at room temperature, affording **12a** in 71% yield although with traces of byproducts; however, 62 h of reaction time was required (Table 1, entry 15). Lowering the

reaction temperature to 60  $^{\circ}$ C reduced the yield of **12a** to 66% (Table 1, entry 16). After a series of attempts, we concluded that the best conversion could be carried out using nitrile (1.0 mmol) and aminoalcohol (5.0 mmol) together with anhydrous Na<sub>2</sub>CO<sub>3</sub> (1.0 mmol) as base for 15 h in dry MeOH (1.0 mL) as solvent under nitrogen atmosphere at 80  $^{\circ}$ C.

The formation of 14 during the reaction of nitrile 11a with 8a led us to speculate 14 as the probable intermediate in the transformation of nitrile 11a to oxazoline 12a. However, reaction with 14 as the starting precursor in place of nitrile 11a did not show any traces of oxazoline formation. This ruled out the participation of 14 as an intermediate in this transformation.

With the optimized reaction conditions, a variety of aryl nitriles produced the desired oxazolines in good to excellent yields (Table 2). Both electron-donating as well as electronwithdrawing groups were well-tolerated. The halide substituted nitriles 11a-d condensed well with 8a, thus, expanding the scope for further derivatization of 2-aryloxazolines. The presence of an electron-donating group such as methyl reduced the reactivity of aryl nitriles in the reaction, affording 74% of 12d as compared to 89% of 12a (Table 2, entries 1 and 4). The reaction with dinitrile, 1,4-dicyanobenzene (11f), produced 71% yield of the corresponding bisoxazoline product 12f selectively with no monooxazoline formation (Table 2, entry 6). Also, the unprotected amino group in nitrile 11g was unaffected under the basic reaction conditions employed in this protocol, thus, producing the corresponding oxazoline 12g in 88% yield (Table 2, entry 7).

The heteroaryl nitriles with a pyridine ring were also found to be compatible substrates under the optimized reaction conditions (Table 3). The 2-heteroaryloxazolines 16a-c were successfully synthesized with nicotinonitrile- and isonicotinonitrile-based substrates in 83–97% yields. Furthermore, the effect of an electron-withdrawing group was noticeable when condensation of 2-chloroisonicotinonitrile (15c) with 8a produced 97% of the desired oxazoline 16c, as compared to isonicotinonitrile (15b) which produced 16b in 85% yield (Table 3, entries 2 and 3).

Furthermore, 2-aryloxazoline derivatives bearing nitrogen heterocycles are also important structural scaffolds in natural product chemistry and pharmaceutical agents that exhibit antidiabetic, anti-Alzheimer's, antibiotic, antihypertensive, and deferrating activities. 1g,h,14 This turned our attention toward the preparation of structurally diverse 2-aryloxazoline derivatives functionalized with synthetically and biologically important Nsubstituted cyclic and acyclic groups such as azoles, ring-fused azoles, saturated heterocycles, and amines (Table 4). As summarized in Table 4, both aliphatic and aromatic heterocycles gave the corresponding 2-aryloxazolines in moderate to good yields (44-71%). The saturated heterocycles such as morpholine, N-acetylated piperazine, piperidine, and pyrrollidine in nitrile precursors 17a-d were well-tolerated in this transformation, affording the corresponding oxazolines 18a-d in moderate to good yields (44-64%, Table 4, entries 1-4). The acetyl group in substrate 17b, which is a reactive site for nucleophilic addition, remained unaffected under the employed reaction conditions. It is worth noting that mGluR5 modulators such as 19 and 20, useful in the treatment of psychiatric and mood disorders, have been partially incorporated in the oxazolines 18c and 18d. 14c Similarly, the indole- and benzimidazole-functionalized oxazoline scaffolds 18g and 18h, which were isolated in 48% and 71% yields respectively (Table

Table 2. Scope of Aryl Nitriles in Oxazoline Formation<sup>a</sup>

	11	8a '`	12
Entry	11	12	Yield (%)
1	Br 11a	N Br 12a	89
2	Br C	12b	83
3	CI 11c	CI 12c	75
4	H <sub>3</sub> C C	Br 12d	74
5	11e	CN O 12e	52
6	NC 11f	N ON N N 12f	71
7	NH <sub>2</sub>	CN NH2 ON N	88

 $^a\mathrm{Reaction}$  conditions: 11 (1.0 mmol), 8a (5.0 mmol), anhydrous  $\mathrm{Na_2CO_3}$  (1.0 mmol), dry MeOH (1.0 mL), 80 °C, 15 h, under nitrogen atmosphere.  $^b\mathrm{Isolated}$  yields.

4, entries 7 and 8), constitute a part of the skeleton of compounds 21 and 22, which are used in the treatment of central nervous system disorders related to or affected by the histamine-3-receptors. Structures of 18g and 18h were confirmed by X-ray diffraction (Figures S1,S2, Supporting Information). Also, the nitriles substituted with N-alkylated or N-benzylated amines 17i-k performed well as precursors in reaction with 8a, affording the desired products 18i-k in 47–69% yields (Table 4, entries 9–11). The NH group that is sensitive to basic conditions was not affected during the reaction of nitrile 17i, thus, providing scope for further derivatization (Table 4, entry 9). However, carbazole bearing nitrile 17l could not be activated toward oxazoline ring formation, which could be due to its poor solubility in methanol.

Table 3. Scope of Heteroaryl Nitriles in Oxazoline Formation  $^a$ 

Entry 15 16 Yield 
$$(\%)^b$$

1 N CN N 83

1 15a 16a

2 N CN N N 85

1 15b 16b

3 CI CN CI N 97

1 15c 16c

<sup>a</sup>Reaction conditions: **15** (1.0 mmol), **8a** (5.0 mmol), anhydrous Na<sub>2</sub>CO<sub>3</sub> (1.0 mmol), dry MeOH (1.0 mL), 80 °C, 15 h, under nitrogen atmosphere. <sup>b</sup>Isolated yields.

After the successful screening of nitriles, a variety of aminoalcohols were screened as the condensation partners in the oxazoline formation reaction. As shown in Table 5, the selected aminoalcohols afforded moderate to excellent yields (28-98%) of the desired products 23a-e under the optimized reaction conditions (Table 5, entries 1-5). Use of 2.0 equiv of Na<sub>2</sub>CO<sub>3</sub> with aminoalcohols 8b and 8c improved the yields of corresponding oxazolines 23a and 23b from 57% to 98% and 55% to 71% respectively (Table 5, entries 1 and 2). However, no significant improvements in the isolated yields of products could be observed with other aminoalcohols 8e and 8f on using 2.0 equiv of Na<sub>2</sub>CO<sub>3</sub>. Moreover, the heteroaryl nitrile 15c proved to be more effective substrate for sterically hindered aminoalcohol 8e, giving oxazoline 23e in 98% yield in contrast to the arvl nitrile 11a which afforded only 28% of compound 23d with 8e (Table 5, entries 4 and 5). The reaction conditions were also suitable for the formation of six-membered oxazine ring in 23f, albeit in moderate (58%) yield (Table 5, entry 6). Additionally, the same conditions could also be applied to the synthesis of 2-arylimidazolines. The use of ethane-1,2-diamine (8g) in place of aminoalcohols 8a-f, produced 2-arylimidazoline 23g in moderate (54%) yield (Table 5, entry 7).

On the basis of above results, a plausible mechanism for the synthesis of oxazolines from nitriles and aminoalcohols is depicted in Scheme 2. The initial step is the in situ formation of imidate I via nucleophilic addition of methanol to nitrile group under basic conditions. Then, I is further attacked by the aminoalcohol nitrogen affording  $\beta$ -hydroxyamidine II which undergoes in situ intramolecular cyclization with subsequent loss of ammonia, to form oxazoline ring (Scheme 2). With reduced amount of aminoalcohol, alcoholysis of the intermediate imidate ester occurs leading to the formation of normal esters via decomposition of ortho esters. Furthermore, the subsequent reaction of normal esters with aminoalcohols produces the transamidation products. Therefore, use of 5.0 mmol of aminoalcohol produced oxazoline 12a as the major

Table 4. Synthesis of Substituted 2-Aryloxazolines $^{a,b}$ 

#### Table 4. continued

"Reaction conditions: 17 (1.0 mmol), 8a (5.0 mmol), anhydrous Na<sub>2</sub>CO<sub>3</sub> (1.0 mmol), dry MeOH (1.0 mL), 80 °C, 15 h, under nitrogen atmosphere. <sup>b</sup>Unreacted nitriles and minor amounts of amides were also isolated along with the desired oxazolines during column purification. <sup>c</sup>Isolated yields. <sup>d</sup>Structure of amide 18ea was confirmed by X-ray diffraction (Figure S3, Supporting Information).

Table 5. Screening of Aminoalcohols a,b

7 11a 
$$H_{2N}$$
  $H_{N}$   $H_{N}$ 

<sup>a</sup>Reaction conditions: 11a/15c (1.0 mmol), 8b-g (5.0 mmol), anhydrous Na<sub>2</sub>CO<sub>3</sub> (1.0 mmol), dry MeOH (1.0 mL), 80 °C, 15 h, under nitrogen atmosphere. <sup>b</sup>Unreacted nitriles and minor amounts of amides were also isolated along with the desired oxazolines during column purification in all the cases excluding entry 5. <sup>c</sup>Isolated yields. <sup>d</sup>Using 2.0 mmol of Na<sub>2</sub>CO<sub>3</sub>.

product, whereas, use of 3.0 mmol of aminoalcohol produced major amount of transamidation product 14 with reduced yield of the oxazoline 12a.

Antioxidant Activity of Synthesized 2-Aryl/Heteroaryloxazolines. The antioxidant activity of the synthesized 2-aryl/heteroaryloxazolines was measured with a simple and rapid DPPH radical scavenging method using ascorbic acid as the

Scheme 2. Proposed Mechanism for the Synthesis of Oxazolines from Nitriles

standard. <sup>16</sup> The sample solutions (1 mg mL<sup>-1</sup>) of all the compounds were prepared in methanol. The freshly prepared DPPH (1.0 mL,  $3 \times 10^{-4}$  M) solution was added to 2.5 mL of the sample solution and allowed to react at room temperature for 30 min. After 30 min, the absorbance was measured at 517 nm. The experiment was repeated three times. The percentage of antioxidant activity are summarized in Table 6. Among all

Table 6. Antioxidant Activity Data for 2-Aryl/Heteroaryloxazolines

compd	DPPH scavenging (%) <sup>a</sup>	compd	DPPH scavenging (%) <sup>a</sup>
12a	28	18f	22
12b	25	18g	52
12c	3	18h	32
12d	32	18i	23
12e	17	18j	40
12f	19	18k	25
12g	35	23a	57
16a	31	23b	22
16b	36	23c	3
16c	49	23d	21
18a	36	23e	16
18b	51	23f	22
18c	25	23g	29
18d	44	ascorbic $acid^b$	97
18e	81		

<sup>a</sup>Results are mean of three different experiments. <sup>b</sup>Standard.

the synthesized 2-aryl/heteroaryloxazolines, 2-(4-((1H-pyrrol1-yl)methyl)phenyl)-4,5-dihydrooxazole (18e) exhibited maximum antioxidant activity (81%) at the concentration of 1 mg mL $^{-1}$  with an IC $_{50}$  value of 320  $\mu g$  mL $^{-1}$  (1.42 mM), whereas compounds 23a, 18b, and 18g showed moderate antioxidant activity (51–57%). Notably, the antioxidant potential of compound 18e could be seen even at lower concentrations (750, 500, 250, and 125  $\mu g$  mL $^{-1}$ ) (Figure 2). The DPPH scavenging potential of 18e was found to decrease with decrease in concentration. The effectiveness of 18e over other compounds in scavenging DPPH activity shows the significance of pyrrole nucleus in enhancing the antioxidant capability of 2-aryloxazolines.

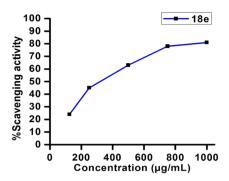


Figure 2. % Scavenging activity vs concentration of compound 18e.

### CONCLUSIONS

In conclusion, a variety of aryl and heteroaryl nitriles were successfully activated under metal- and catalyst-free conditions delivering 2-aryl/heteroaryloxazoline scaffolds with wide skeletal diversity. A small library of 2-aryloxazolines functionalized with biologically active motifs such as azoles, ring-fused azoles, saturated heterocyclics, and amines could be successfully synthesized in moderate to good yields. The pyrrole-functionalized 2-aryloxazoline displayed maximum antioxidant activity among all the 2-aryl/heteroaryloxazolines tested. The general applicability of the experimental procedure, operational simplicity, ready availability of the starting materials, and presence of functional group handle in many of the synthesized 2-aryl/heteroaryloxazolines will promote their synthesis for pharmaceutical applications.

#### **■ EXPERIMENTAL SECTION**

**General Information.** Unless otherwise noted, all reactions were carried out under nitrogen atmosphere using standard Schlenk techniques. All solvents were dried according to standard procedures. Column chromatography was carried out with silica gel 60-120 mesh. The  $^1H$  (400 MHz) and  $^{13}C\{^1H\}$  (100 MHz) NMR spectra were recorded in CDCl<sub>3</sub> at 25 °C. The chemical shifts are expressed in parts per million ( $\delta$ ) relative to residual solvent protons of CDCl<sub>3</sub>:  $\delta$  7.24 for  $^1H$  NMR and  $\delta$  77.0 for  $^{13}C\{^1H\}$  NMR. The following abbreviations are used for multiplicities: br = broad, s = singlet, d = doublet, t = triplet, q = quartet, and m = multiplet. High-resolution mass spectra were measured on a high-resolution TOF instrument with electrospray ionization (ESI). Melting points are uncorrected. The nitrile substrates 17a,  $^{18}$  17c,  $^{19}$  and 17d– $1^{20-27}$  were prepared according to the literature.

Synthesis of 4-(2-Aminoethoxy)methyl)benzonitrile (11g). A mixture of 4-(bromomethyl)benzonitrile (0.981g, 5.0 mmol), 2-aminoethanol (1.53 g, 25.0 mmol), and anhydrous Na<sub>2</sub>CO<sub>3</sub> (0.53 g, 5.0 mmol) in dry MeOH (5.0 mL) was stirred under nitrogen atmosphere at 80 °C for 4 h. On completion of reaction, the reaction mixture was concentrated to give a viscous residue. Purification of the crude reaction mixture by column chromatography on silica gel (CHCl<sub>3</sub>/MeOH = 32:1 afforded compound 11g (0.705 g, 80%) as a yellow solid. mp = 71–73 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.59 (d, J = 8.2 Hz, 2H), 7.42 (d, J = 8.3 Hz, 2H), 3.85 (s, 2H), 3.66 (t, J = 5.0 Hz, 2H), 2.77 (t, J = 5.1 Hz, 2H), 1.93 (br s, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 145.3, 132.1, 128.6, 118.8, 110.6, 60.7, 52.9, 50.5; IR (KBr, v cm $^{-1}$ ): 3422, 3246, 3089, 2920, 2861, 2228, 1625, 1492, 1347, 1295, 1235, 1204, 1179, 1143, 1106,

1053, 990, 961, 923, 856; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>10</sub>H<sub>13</sub>N<sub>2</sub>O, 177.1022; found, 177.1026.

Synthesis of 4-((4-Acetylpiperazin-1-yl)methyl)benzonitrile (17b). A mixture of 4-(bromomethyl)benzonitrile (0.981 g, 5.0 mmol), N-acetylpiperazine (0.641 g, 5.0 mmol), and potassium hydroxide (1.12 g, 20.0 mmol) in dry THF (10.0 mL) was stirred under nitrogen atmosphere at room temperature for 2 days. On completion of reaction, the reaction mixture was concentrated to give a viscous residue. Purification of the crude reaction mixture by column chromatography on silica gel (CHCl<sub>3</sub>/MeOH = 32:1 afforded compound 17b (1.075 g, 88%) as a white solid. mp =  $68-70 \, ^{\circ}\text{C}$ ; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.60 (d, J = 8.1 Hz, 2H), 7.43 (d, J = 8.1 Hz, 2H), 3.60 (t, I = 5.1 Hz, 2H), 3.54 (s, 2H), 3.44 (t, I = 5.1 Hz, 2H),2.42-2.38 (m, 4H), 2.06 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  168.9, 143.4, 132.2, 129.4, 118.8, 111.1, 62.2, 53.1, 52.7, 46.2, 41.3, 21.3; IR (KBr,  $v \text{ cm}^{-1}$ ): 3462, 3068, 2997, 2919, 2867, 2810, 2225, 1638, 1438, 1421, 1368, 1350, 1306, 1250, 1138, 995; HRMS (ESI-TOF) m/z:  $[M + H]^+$  calcd for C<sub>14</sub>H<sub>18</sub>N<sub>3</sub>O, 244.1444; found, 244.1451.

General Experimental Procedure for the Synthesis of 2-Aryl/Heteroaryloxazolines. To a solution of nitrile (1.0 mmol) and anhydrous  $Na_2CO_3$  (106 mg, 1.0 mmol) in dry methanol (1.0 mL) under nitrogen atmosphere, aminoalcohol (5.0 mmol) was added, and the reaction mixture was stirred at 80 °C for 15 h. After 15 h, the reaction mixture was cooled to room temperature and concentrated. Purification of the crude reaction mixture by column chromatography on silica gel (PE/EtOAc or CHCl<sub>3</sub>/MeOH) afforded the desired 2-aryl/heteroaryloxazoline products.

2-(4-Bromophenyl)-4,5-dihydrooxazole (12a). <sup>12j</sup> Column chromatography purification (PE/EtOAc = 9:1) afforded compound 12a (198.2 mg, 89%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.76 (d, J = 8.2 Hz, 2H), 7.50 (d, J = 8.3 Hz, 2H), 4.38 (t, J = 9.6 Hz, 2H), 4.00 (t, J = 9.6 Hz, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.8, 131.5, 129.6, 126.7, 125.9, 67.7, 54.9.

2-(3-Bromophenyl)-4,5-dihydrooxazole (12b). <sup>91</sup> Column chromatography purification (PE/EtOAc = 9:1) afforded compound 12b (187.8 mg, 83%) as a colorless viscous liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.02 (s, 1H), 7.79 (d, J = 7.9 Hz, 1H), 7.52 (d, J = 8.0 Hz, 1H), 7.20 (t, J = 7.9 Hz, 1H), 4.36 (t, J = 9.8 Hz, 2H), 3.99 (t, J = 9.5 Hz, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.2, 134.1, 131.0, 129.8, 129.5, 126.6, 122.2, 67.7, 54.8.

2-(4-Chlorophenyl)-4,5-dihydrooxazole (12c). <sup>12j</sup> Column chromatography purification (PE/EtOAc = 9:1) afforded compound 12c (135.6 mg, 75%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.86 (d, J = 8.5 Hz, 2H), 7.36 (d, J = 8.6 Hz, 2H), 4.42 (t, J = 9.8 Hz, 2H), 4.04 (t, J = 9.8 Hz, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.7, 137.3, 129.4, 128.5, 126.2, 67.7, 54.9.

2-(4-Bromo-3-methylphenyl)-4,5-dihydrooxazole (12d). Column chromatography purification (PE/EtOAc = 19:1) afforded compound 12d (178.3 mg, 74%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.77 (s, 1H), 7.56–7.50 (m, 2H), 4.37 (t, J = 9.6 Hz, 2H), 3.99 (t, J = 9.6 Hz, 2H), 2.37 (s, 3H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.9, 138.0, 132.3, 130.2, 128.3, 126.8, 126.7, 67.6, 54.8, 22.7.

2-((1,1'-Biphenyl)-4-yl)-4,5-dihydrooxazole (12e).<sup>10</sup> Column chromatography purification (PE/EtOAc = 9:1) afforded compound 12e (115.8 mg, 52%) as a white solid. <sup>1</sup>H NMR

(400 MHz, CDCl<sub>3</sub>):  $\delta$  8.03 (d, J = 8.5 Hz, 2H), 7.65–7.60 (m, 4H), 7.46–7.42 (m, 2H), 7.38–7.34 (m, 1H), 4.48 (t, J = 9.8 Hz, 2H), 4.09 (t, J = 9.8 Hz, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.5, 143.9, 140.2, 128.8, 128.6,127.9, 127.1, 127.0, 126.5, 67.6, 54.9.

1,4-Bis(4,5-dihydrooxazol-2-yl)benzene (12f). Column chromatography purification (PE/EtOAc = 1:1) afforded compound 12f (152.5 mg, 71%) as a white solid. H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.96 (s, 4H), 4.42 (t, J = 9.8 Hz, 4H), 4.05 (t, J = 9.8 Hz, 4H); NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.0, 130.2, 128.1, 67.7, 55.0.

2-((4-(4,5-Dihydrooxazol-2-yl)benzyl)oxy)ethanamine (12g). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 13:1) afforded compound 12g (194.3 mg, 88%) as a white solid. mp = 49–51 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.89 (d, J = 8.2 Hz, 2H), 7.38 (d, J = 8.2 Hz, 2H), 4.41 (t, J = 9.6 Hz, 2H), 4.03 (t, J = 9.6 Hz, 2H), 3.87 (s, 2H), 3.67 (t, J = 5.1 Hz, 2H), 2.81 (t, J = 5.0 Hz, 2H), 2.66 (br s, 2H);  ${}^{13}$ C{ ${}^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.5, 143.1, 128.3, 128.1, 126.6, 67.6, 60.8, 54.9, 53.1, 50.5; IR (KBr, v cm ${}^{-1}$ ): 3312, 2934, 2850, 1647, 1572, 1513, 1458, 1417, 1364, 1262, 1217, 1076, 1020, 978, 945, 838, 769, 682, 667; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>12</sub>H<sub>17</sub>N<sub>2</sub>O<sub>2</sub>, 221.1285; found, 221.1295.

2-(Pyridin-3-yl)-4,5-dihydrooxazole (16a). <sup>12j</sup> Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 16a (123.1 mg, 83%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 9.11 (d, J = 1.8 Hz, 1H), 8.66 (dd, J = 5.0, 1.8 Hz, 1H), 8.17 (dt, J = 7.8, 1.8 Hz, 1H), 7.31 (ddd, J = 7.8, 5.0, 0.9 Hz, 1H), 4.42 (t, J = 9.6 Hz, 2H), 4.04 (t, J = 9.6 Hz, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 162.6, 151.9, 149.4, 135.4, 123.8, 123.1, 67.7, 54.9.

2-(Pyridin-4-yl)-4,5-dihydrooxazole (16b). <sup>12j</sup> Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 16b (126 mg, 85%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.69 (d, J = 5.5 Hz, 2H), 7.77 (d, J = 6.1 Hz, 2H), 4.45 (t, J = 9.8 Hz, 2H), 4.08 (t, J = 9.8 Hz, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.0, 150.2, 135.1, 121.9, 67.9, 55.1.

2-(2-Chloropyridin-4-yl)-4,5-dihydrooxazole (16c). Column chromatography purification (PE/EtOAc = 4:1) afforded compound 16c (177.5 mg, 97%) as a white solid. mp = 102–103 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.45 (d, J = 4.9 Hz, 1H), 7.80 (s, 1H), 7.70–7.69 (m, 1H), 4.46 (t, J = 9.8 Hz, 2H), 4.09 (t, J = 9.8 Hz, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 161.9, 152.1, 150.1, 138.0, 122.8, 120.6, 68.1, 55.1; IR (KBr, v cm<sup>-1</sup>): 3313, 2925, 1653, 1595, 1545, 1473, 1388, 1259, 1137, 1077, 976, 942, 775, 728, 696, 476; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>8</sub>H<sub>8</sub>ClN<sub>2</sub>O, 183.0320; found, 183.0350.

2-(4-(Morpholin-4-ylmethyl)phenyl)-4,5-dihydrooxazole (18a). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 18a (125 mg, 51%) as a light yellow solid. mp = 67–69 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.85 (d, J = 8.2 Hz, 2H), 7.34 (d, J = 8.3 Hz, 2H), 4.38 (t, J = 9.6 Hz, 2H), 4.01 (t, J = 9.6 Hz, 2H), 3.68–3.66 (m, 4H), 3.49 (s, 2H), 2.42–2.39 (m, 4H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.5, 141.2, 129.0, 128.1, 126.6, 67.5, 66.8, 63.0, 54.8, 53.5; IR (KBr, v cm $^{-1}$ ): 2957, 2855, 2810, 1718, 1651, 1573, 1513, 1480, 1455, 1417, 1351, 1333, 1286, 1261, 1116, 1071, 1035, 1008, 976, 944, 915, 867, 799, 756, 680; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>14</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub>, 247.1441; found, 247.1443.

1-(4-(4-(4,5-Dihydrooxazol-2-yl)benzyl)piperazin-1-yl)-ethanone (18b). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 18b (127.3 mg,

44%) as a white solid. mp = 144.145 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.87 (d, J = 8.2 Hz, 2H), 7.34 (d, J = 8.2 Hz, 2H), 4.40 (t, J = 9.6 Hz, 2H), 4.03 (t, J = 9.6 Hz, 2H), 3.59 (t, J = 5.0 Hz, 2H), 3.53 (s, 2H), 3.43 (t, J = 5.0 Hz, 2H), 2.41–2.38 (m, 4H), 2.04 (s, 3H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  168.9, 164.4, 141.1, 128.9, 128.1, 126.7, 67.5, 62.4, 54.8, 53.0, 52.7, 46.2, 41.3, 21.3; IR (KBr, v cm $^{-1}$ ): 2943, 2908, 2804, 2771, 1629, 1434, 1349, 1309, 1268, 1256, 1235, 1145, 1104, 1068, 1017, 1002, 988, 940, 898, 850, 732, 676, 622, 582; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>16</sub>H<sub>22</sub>N<sub>3</sub>O<sub>2</sub>, 288.1707; found, 288.1715.

2-(4-(Piperidin-1-ylmethyl)phenyl)-4,5-dihydrooxazole (18c). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 18c (155.6 mg, 64%) as a light yellow solid. mp = 70–72 °C;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.86 (d, J = 8.6 Hz, 2H), 7.36 (d, J = 8.6 Hz, 2H), 4.39 (t, J = 9.5 Hz, 2H), 4.02 (t, J = 9.5 Hz, 2H), 3.52 (s, 2H), 2.39 (br s, 4H), 1.61–1.55 (m, 4H), 1.43–1.40 (m, 2H);  $^{13}$ C( $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.6, 129.3, 128.1, 126.6, 67.5, 63.2, 54.9, 54.3, 25.6, 24.0; IR (KBr, v cm $^{-1}$ ): 3281, 2930, 2854, 2795, 1718, 1652, 1458, 1417, 1362, 1261, 1101, 1069, 1039, 1020, 945, 865, 801, 679; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>1</sub>cH<sub>2</sub>1N<sub>2</sub>O, 245.1648; found, 245.1657.

2-(4-(Pyrrolidin-1-ylmethyl)phenyl)-4,5-dihydrooxazole (18d). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 9:1) afforded compound 18d (130.3 mg, 57%) as a yellow viscous liquid.  $^1$ H NMR (400 MHz, CDCl<sub>3</sub> δ 7.83 (d, J = 8.2 Hz, 2H), 7.32 (d, J = 8.2 Hz, 2H), 4.36 (t, J = 9.6 Hz, 2H), 3.99 (t, J = 9.6 Hz, 2H), 3.59 (s, 2H), 2.46–2.45 (m, 4H), 1.74–1.72 (m, 4H);  $^{13}$ C{ $^1$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.6, 142.7, 128.7, 128.0, 126.2, 67.5, 60.3, 54.8, 54.1, 23.3; IR (KBr, v cm $^{-1}$ ): 2966, 2877, 2797, 1648, 1416, 1363, 1330, 1261, 1196, 1125, 1070, 1020, 976, 944, 881, 849, 730, 681; HRMS (ESI-TOF) m/z: [M + H] $^+$  calcd for C<sub>14</sub>H<sub>19</sub>N<sub>2</sub>O, 231.1492; found, 231.1505.

2-(4-((1H-Pyrrol-1-yl)methyl)phenyl)-4,5-dihydrooxazole (18e). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 99:1) afforded compound 18e (103.7 mg, 46%) as a white solid. mp = 75–76 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.89 (d, J = 8.3 Hz, 2H), 7.12 (d, J = 8.2 Hz, 2H), 6.68–6.67 (m, 2H), 6.19–6.18 (m, 2H), 5.10 (s, 2H), 4.42 (t, J = 9.6 Hz, 2H), 4.04 (t, J = 9.6 Hz, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.2, 141.4, 128.5, 127.0, 126.7, 121.1, 108.7, 67.5, 54.8, 52.9; IR (KBr, v cm $^{-1}$ ): 2928, 1641, 1390, 1261, 1182, 1068, 940, 862, 732, 696; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>14</sub>H<sub>15</sub>N<sub>2</sub>O, 227.1179; found, 227.1172.

2-(4-(2-Methylimidazol-1-ylmethyl)-phenyl)-4,5-dihydrooxazole (18f). Purification by preparative TLC on silica gel G (CHCl<sub>3</sub>/MeOH = 19:1) afforded compound 18f (144.2 mg, 60%) as a dark yellow viscous liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.80 (d, J = 8.2 Hz, 2H), 6.97 (d, J = 8.2 Hz, 2H), 6.85 (s, 1H), 6.75 (s, 1H), 4.97 (s, 2H), 4.31 (t, J = 9.6 Hz, 2H), 3.93 (t, J = 9.6 Hz, 2H), 2.19 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.7, 144.6, 139.3, 128.4, 127.1, 127.0, 126.2, 119.7, 67.4, 54.5, 49.1, 12.6; IR (KBr, v cm<sup>-1</sup>): 2923, 1648, 1420, 1364, 1263, 1078, 1020, 987, 943, 832, 724, 688; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>16</sub>N<sub>3</sub>O, 242.1288; found, 242.1292.

1-(4-(4,5-Dihydrooxazol-2-yl)-benzyl)-1H-indole (18g). Column chromatography purification (PE/EtOAc = 9:1) afforded compound 18g (133.3 mg, 48%) as a yellow solid. mp = 122–123 °C;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.87 (d, J = 8.2 Hz, 2H), 7.66 (d, J = 8.2 Hz, 1H), 7.26–7.22 (m, 1H),

7.18–7.10 (m, SH), 6.58–6.57 (m, 1H), 5.36 (s, 2H), 4.41 (t, J = 9.6 Hz, 2H), 4.04 (t, J = 9.6 Hz, 2H);  $^{13}$ C $^{1}$ H $^{13}$ NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.2, 140.8, 136.1, 128.7, 128.6, 128.2, 127.0, 126.6, 121.8, 121.0, 119.6, 109.6, 101.9, 67.6, 54.8, 49.8; IR (KBr, v cm<sup>-1</sup>): 3046, 2929, 2874, 1648, 1459, 1418, 1356, 1333, 1316, 1260, 1193, 1179, 1068, 1016, 976, 943, 824, 748, 722, 671; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for  $C_{18}H_{17}N_2O$ , 277.1335; found, 277.1343.

2-(4-((1H-Benzo[d]imidazol-1-yl)methyl)phenyl)-4,5-dihydrooxazole (18h). Column chromatography purification (PE/EtOAc = 9:1) afforded compound 18h (196.9 mg, 71%) as a white solid. mp = 154–155 °C; ¹H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.94 (s, 1H), 7.90–7.87 (m, 2H), 7.82–7.80 (m, 1H), 7.28–7.17 (m, 5H), 5.37 (s, 2H), 4.39 (t, J = 9.6 Hz, 2H), 4.02 (t, J = 9.6 Hz, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.0, 143.9, 143.1, 138.6, 133.7, 128.8, 127.8, 126.9, 123.2, 122.4, 120.4, 109.9, 67.6, 54.9, 48.5; IR (KBr, v cm $^{-1}$ ): 2927, 2903, 2874, 1648, 1611, 1497, 1460, 1434, 1415, 1359, 1286, 1261, 1200, 1188, 1066, 1014, 972, 942, 826, 752, 721, 670; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>17</sub>H<sub>16</sub>N<sub>3</sub>O, 278.1288; found, 278.1294.

*N-*(*4-*(*4,5-Dihydrooxazol-2-yl)benzyl)butan-1-amine* (*18i*). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 13:1) afforded compound *18i* (135.9 mg, 59%) as a yellow viscous liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.82 (d, *J* = 8.5 Hz, 2H), 7.29 (d, *J* = 8.6 Hz, 2H), 4.34 (t, *J* = 9.8 Hz, 2H), 3.96 (t, *J* = 9.8 Hz, 2H), 3.74 (s, 2H), 2.54 (t, *J* = 7.3 Hz, 2H), 2.27 (br s, 1H), 1.46–1.38 (m, 2H), 1.31–1.22 (m, 2H), 0.83 (t, *J* = 7.3 Hz, 3H); <sup>13</sup>C{ <sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.4, 143.7, 128.1, 127.8, 126.1, 67.4, 54.6, 53.4, 48.9, 31.9, 20.3, 13.8; IR (KBr, v cm<sup>-1</sup>): 3292, 2960, 2875, 2485, 1925, 1648, 1613, 1573, 1513, 1458, 1416, 1362, 1262, 1216, 1074, 1106, 1074, 1020, 977, 945, 850, 753, 680, 665; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>21</sub>N<sub>2</sub>O, 233.1648; found, 233.1655.

*N-Benzyl-(4-(4,5-dihydrooxazol-2-yl)benzyl)methylamine* (*18j*). Column chromatography purification (PE/EtOAc = 19:1) afforded compound **18j** (192 mg, 69%) as a white solid. mp = 54–56 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.84 (d, J = 8.3 Hz, 2H), 7.36 (d, J = 8.2 Hz, 2H), 7.31–7.24 (m, 4H), 7.21–7.17 (m, 1H), 4.36 (t, J = 9.6 Hz, 2H), 3.99 (t, J = 9.4 Hz, 2H), 3.51–3.49 (m, 4H), 2.13 (s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.6, 142.8, 138.9, 128.9, 128.8, 128.2, 128.0, 127.0, 126.3, 67.5, 61.8, 61.4, 54.8, 42.2; IR (KBr, v cm<sup>-1</sup>): 3029, 2934, 2840, 2788, 1649, 1452, 1413, 1361, 1260, 1070, 1019, 975, 944, 871, 846, 736, 699, 679; HRMS (ESI-TOF) m/z:  $[M + H]^+$  calcd for  $C_{18}H_{21}N_2O$ , 281.1648; found, 281.1643.

N,N-Dibenzyl-1-(4-(4,5-dihydrooxazol-2-yl)phenyl)-methanamine (18k). Column chromatography purification (PE/EtOAc = 2:1) afforded compound 18k (137.6 mg, 47%) as a colorless viscous liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.82 (d, J = 8.6 Hz, 2H), 7.38 (d, J = 8.6 Hz, 2H), 7.31 (d, J = 7.3 Hz, 4H), 7.24 (t, J = 7.3 Hz, 4H), 7.17–7.13 (m, 2H), 4.33 (t, J = 9.8 Hz, 2H), 3.96 (t, J = 9.8 Hz, 2H), 3.50 (s, 2H), 3.47 (s, 4H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 164.6, 143.3, 139.3, 128.7, 128.6(8), 128.2, 128.1, 127.0, 126.3, 67.5, 57.9, 57.6, 54.8; IR (KBr, v cm<sup>-1</sup>): 3061, 3028, 2929, 2878, 2799, 1649, 1494, 1453, 1416, 1362, 1329, 1260, 1122, 1070, 1020, 975, 944, 746, 699, 680; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>24</sub>H<sub>25</sub>N<sub>2</sub>O, 357.1961; found, 357.1977.

4-((1H-Pyrrol-1-yl)methyl)benzamide (18ea). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 49:1) afforded compound 18ea (30 mg, 15%) as a white solid. mp = 105-106 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.75 (d, J = 8.8 Hz, 2H),

7.14 (d, J = 8.7 Hz, 2H), 6.68 (t, J = 2.0 Hz, 2H), 6.20 (t, J = 2.0 Hz, 2H), 6.11 (br, 1H), 5.95 (br, 1H), 5.11 (s, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  169.0, 142.5, 127.8, 127.4, 127.0, 121.2, 108.9, 52.9; IR (film, v cm $^{-1}$ ): 3386, 3185, 2923, 2853, 1648, 1396, 1282, 1087, 723; HRMS (ESI-TOF) m/z: [M + Na] $^{+}$  calcd for C $_{12}$ H $_{12}$ N $_{2}$ ONa, 223.0842; found, 223.0846.

2-(4-Bromophenyl)-4,5-dihydro-4-methyloxazole (23a). Column chromatography purification (PE/EtOAc = 9:1) afforded compound 23a (135.9 mg, 57%) as a colorless liquid. With 2.0 mmol of Na<sub>2</sub>CO<sub>3</sub>, 98% (234.6 mg) of 23a was isolated. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.79 (d, J = 8.5 Hz, 2H), 7.52 (d, J = 8.5 Hz, 2H), 4.53-4.49 (m, 1H), 4.40-4.31 (m, 1H), 3.94 (t, J = 7.9 Hz, 1H), 1.34 (d, J = 6.1 Hz, 3H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 162.7, 131.6, 129.7, 126.7, 125.9, 74.2, 62.0, 21.3; IR (KBr, v cm $^{-1}$ ): 3284, 2973, 2930, 2871, 1718, 1648, 1593, 1488, 1452, 1399, 1366, 1333, 1260, 1174, 1077, 1012, 1174, 1077, 1012, 990, 887, 837, 727, 670; HRMS (ESI-TOF) m/z: [M + H] $^{+}$  calcd for C<sub>10</sub>H<sub>11</sub>BrNO, 240.0019; found, 240.0025; [ $\alpha$ ]<sub>D</sub><sup>28</sup> +57.4 (c 1.0 in CHCl<sub>3</sub>).

2-(4-Bromophenyl)-4,5-dihydro-5-methyloxazole (23b). Column chromatography purification (PE/EtOAc = 9:1) afforded compound 23b (131.8 mg, 55%) as a colorless liquid. With 2.0 mmol of Na<sub>2</sub>CO<sub>3</sub>, 71% (170.7 mg) of 23b was isolated. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.77 (d, J = 8.5 Hz, 2H), 7.50 (d, J = 8.6 Hz, 2H), 4.86–4.77 (m, 1H), 4.10 (dd, J = 14.7, 9.8 Hz, 1H), 3.56 (dd, J = 14.7, 7.9 Hz, 1H), 1.39 (d, J = 6.1 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 163.0, 131.5, 129.6, 126.9, 125.8, 76.5, 61.6, 21.0; IR (KBr, v cm<sup>-1</sup>): 3284, 2973, 2930, 2871, 1718, 1648, 1593, 1488, 1452, 1399, 1366, 1333, 1260, 1174, 1077, 1012, 990, 887, 837, 727, 670; HRMS (ESI-TOF) m/z: [M + H]<sup>+</sup> calcd for C<sub>10</sub>H<sub>11</sub>BrNO, 240.0019; found, 240.0021; [ $\alpha$ ]<sub>D</sub><sup>28</sup> −18.0 (c 1.0 in CHCl<sub>3</sub>).

2-(4-Bromophenyl)-4,5-dihydro-4-hydroxymethyl-4-methyloxazole (23c). Column chromatography purification (CHCl<sub>3</sub>/MeOH = 99:1) afforded compound 23c (247.1 mg, 92%) as a white solid. H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.67 (d, J = 8.7 Hz, 2H), 7.45 (d, J = 8.7 Hz, 2H), 4.48 (d, J = 8.2 Hz, 1H), 4.07 (d, J = 8.2 Hz, 1H), 3.77 (d, J = 11.9 Hz, 1H), 3.45 (d, J = 11.9 Hz, 1H), 3.06 (br, 1H), 1.29 (s, 3H);  ${}^{13}$ C{ ${}^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>): δ 163.6, 131.5, 129.9, 126.4, 125.9, 75.0, 72.0, 67.8, 23.6.

2-(4-Bromophenyl)-4,5-dihydro-4,4-dimethyloxazole (23d). <sup>9c</sup> Column chromatography purification (PE/EtOAc = 19:1) afforded compound 23d (70.4 mg, 28%) as a colorless liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.77 (d, J = 8.6 Hz, 2H), 7.51 (d, J = 8.6 Hz, 2H), 4.08 (s, 2H), 1.35 (s, 6H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  161.3, 131.5, 129.7, 126.9, 125.8, 79.2, 67.7, 28.3.

2-(2-Chloropyridin-4-yl)-4,5-dihydro-4,4-dimethyloxazole (23e). Column chromatography purification (PE/EtOAc = 19:1) afforded compound 23e (205.8 mg, 98%) as a colorless liquid. H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.40–8.38 (m, 1H), 7.76 (s, 1H), 7.64–7.62 (m, 1H), 4.09 (s, 2H), 1.32 (s, 6H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  159.1, 151.9, 149.9, 138.3, 122.8, 120.6, 79.4, 68.1, 28.1.

2-(4-Bromophenyl)-5,6-dihydro-4H-1,3-oxazine (23f). <sup>12j</sup> Column chromatography purification (PE/EtOAc = 9:1) afforded compound 23f (138.2 mg, 58%) as a colorless liquid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.71 (d, J = 8.5 Hz, 2H), 7.44 (d, J = 8.6 Hz, 2H), 4.29 (t, J = 5.5 Hz, 2H), 3.53 (t, J = 5.8 Hz,

2H), 1.95–1.89 (m, 2H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  154.7, 132.9, 131.0, 128.4, 124.7, 65.1, 42.5, 21.7.

2-(4-Bromophenyl)-4,5-dihydro-1H-imidazole (23g).<sup>31</sup> Column chromatography purification (CHCl<sub>3</sub>/MeOH = 16:1) afforded compound 23g (120.8 mg, 54%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.64 (d, J = 8.5 Hz, 2H), 7.52 (d, J = 8.6 Hz, 2H), 3.77 (s, 4H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.8, 131.6, 129.3, 128.5, 125.0, 50.4.

*4-Bromobenzamide* (13). Column chromatography purification (PE/EtOAc = 4:1) afforded compound 13 (18 mg, 9%) as a white solid (Table 1, entry 1). H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.03 (br s, 1H), 7.80 (d, J = 8.8 Hz, 2H), 7.65 (d, J = 8.1 Hz, 2H), 7.45 (br s, 1H);  $^{13}$ C{ $^{1}$ H} NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  166.9, 133.4, 131.2, 129.6, 125.0.

4-Bromo-N-(2-hydroxyethyl)benzamide (14).<sup>33</sup> Column chromatography purification (EtOAc) afforded compound 14 (107.4 mg, 44%) as a white solid (Table 1, entry 1). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 8.52 (t, J = 5.9 Hz, 1H), 7.79 (d, J = 8.8 Hz, 2H), 7.66 (d, J = 8.1 Hz, 2H), 4.72 (t, J = 5.5 Hz, 1H), 3.49 (q, J = 5.9 Hz, 2H), 3.31–3.28 (m, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, DMSO- $d_6$ ): δ 165.4, 133.5, 131.2, 129.3, 124.6, 59.5, 42.2.

#### ASSOCIATED CONTENT

# **S** Supporting Information

Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compounds **11g**, **17b**, **12a**–**g**, **16a**–**c**, **18a**–**k**, **18ea**, **23a**–**g**, **13**, and **14**, HRMS spectra for **11g**, **17b**, **12g**, **16c**, **18a**–**k**, **18ea**, **23a** and **23b** along with CIF of **18g**, **18h** and **18ea** are available. This material is available free of charge via the Internet at http://pubs.acs.org.

# AUTHOR INFORMATION

# **Corresponding Author**

\*E-mail: mdmilton@chemistry.du.ac.in.

#### Notes

The authors declare no competing financial interest.

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# DEDICATION

Dedicated to the memory of Dr. Anita Tandon.

### REFERENCES

(1) (a) Tsuda, M.; Yamakawa, M.; Oka, S.; Tanaka, Y.; Hoshino, Y.; Mikami, Y.; Sato, A.; Fujiwara, H.; Ohizumi, Y.; Kobayashi, J. J. Nat. Prod. 2005, 68, 462. (b) Onishi, H. R.; Pelak, B. A.; Silver, L. L.; Kahan, F. M.; Chen, M.-H.; Patchett, A. A.; Galloway, S. M.; Hyland, S. A.; Anderson, M. S.; Raetz, C. R. H. Science 1996, 274, 980. (c) Li, Q.; Woods, K. W.; Claireborne, A.; Gwanltey, S. L., II; Barr, K. J.; Liu, G.; Gehrke, L.; Credo, R. B.; Hua Hui, Y.; Lee, J.; Warner, R. B.; Kovar, P.; Nukkla, M. A.; Zielinski, N. A.; Tahir, S. K.; Fitzgerald, M.; Kim, K. H.; Marsh, K.; Frost, D.; Ng, S.-C.; Rosenberg, S.; Sham, H. L. Bioorg. Med. Chem. 2002, 12, 465. (d) Tsukamoto, M.; Murooka, K.; Nakajima, S.; Abe, S.; Suzuki, H.; Hirano, K.; Kondo, H.; Kojiri, K.; Suda, H. J. Antibiot. 1997, 50, 815. (e) Vizi, E. S. Med. Res. Rev. 1986, 6, 431. (f) Celanire, S.; Talaga, P.; Leurs, R.; Denonne, F.; Timmerman, H.; Lebon, F. Patent no. WO 2006103057 A1, 2006.

- (g) Gross, J. L.; Robichaud, A. J.; Mazzacani, A.; Williams, M. J. Patent no. US 0023707, 2009. (h) Bergeron, R. J.; Xin, M. G.; Weimar, W. R.; Smith, R. E.; Wiegand, J. *J. Med. Chem.* **2001**, 44, 2469.
- (2) For representative reviews, see: (a) Gant, T. G.; Meyers, A. I. Tetrahedron 1994, 50, 2297. (b) Reuman, M.; Meyers, A. I. Tetrahedron 1985, 41, 837. (c) Frump, J. A. Chem. Rev. 1971, 71, 483. (3) (a) Lee, Y.-J.; Lee, J.; Kim, M.-J.; Jeong, B.-S.; Lee, J.-H.; Kim, T.-
- S.; Lee, J.; Ku, J.-M.; Jew, S.-S.; Park, H.-G. Org. Lett. 2005, 7, 3207. (b) Saravanan, P.; Corey, E. J. J. Org. Chem. 2003, 68, 2760. (c) Greene, T. W.; Wutz, P. G. M. Protective Groups in Organic Synthesis, 2nd ed; John Wiley & Sons: New York, 1991.
- (4) Hargaden, G. C.; Guiry, P. J. Chem. Rev. 2009, 109, 2505.
- (5) For some selected examples, see: (a) Khumsubdee, S.; Fan, Y.; Burgess, K. J. Org. Chem. 2013, 78, 9969. (b) Mckeon, S. C.; Muller-Bunz, H.; Guiry, P. J. Eur. J. Org. Chem. 2011, 7107. (c) Sawada, T.; Nakada, M. Adv. Synth. Catal. 2005, 347, 1527.
- (6) Adams, N.; Úlrich, S. Adv. Drug Delivery Rev. 2007, 59, 1504.
- (7) (a) Kuwano, R.; Kameyama, N.; Ikeda, R. *J. Am. Chem. Soc.* **2011**, 133, 7312. (b) Ramirez, T. A.; Zhao, B.; Shi, Y. *Tetrahedron Lett.* **2010**, 51, 1822. (c) Lui, L.; Floreancig, P. E. *Org. Lett.* **2010**, 12, 4686. (d) Baba, D.; Fuchigami, T. *Tetrahedron Lett.* **2003**, 44, 3133.
- (8) (a) Padmaja, A.; Rajasekhar, C.; Durgamma, S.; Venkatesh, B. C.; Padmavathi, V. Med. Chem. Res. 2014, 23, 1084. (b) Djurendic, E.; Vujaskovic, S. D.; Sakac, M.; Ajdukovic, J.; Gakovic, A.; Kojic, V.; Bogdanovic, G.; Klisuric, O.; Gasi, G. P. ARKIVOC 2011, (ii), 83. (c) Padmavathi, V.; Mahesh, K.; Reddy, G. D.; Padmaja, A. Eur. J. Med. Chem. 2010, 45, 3178.
- (9) For selected examples, see: (a) Boissarie, P. J.; Hamilton, Z. E.; Lang, S.; Murphy, J. A.; Suckling, C. J. Org. Lett. 2011, 13, 6256. (b) Ilkgul, B.; Gunes, D.; Sirkecioglu, O.; Bicak, N. Tetrahedron Lett. 2010, 51, 5313. (c) Kangani, C. O.; Day, B. W. Tetrahedron Lett. 2009, 50, 5332. (d) Takahashi, S.; Togo, H. Synthesis 2009, 2329. (e) Clayden, J.; Clayton, J.; Harvey, R. A.; Karlubikova, O. Synlett 2009, 2836. (f) Kempe, K.; Lobert, M.; Hoogenboom, R.; Schubert, U. S. J. Comb. Chem. 2009, 11, 274. (g) Minakata, S.; Morino, Y.; Ide, T.; Oderaotoshi, Y.; Komatsu, M. Chem. Commun. 2007, 3279. (h) Ishihara, M.; Togo, H. Tetrahedron 2007, 63, 1474. (i) Sayama, S. Synlett 2006, 1479. (j) Cwik, A.; Hell, Z.; Hegedus, A.; Finta, Z.; Horvath, Z. Tetrahedron Lett. 2002, 43, 3985. (k) Wipf, P.; Wang, X. J. Comb. Chem. 2002, 4, 656.
- (10) Wu, X.-F.; Neumann, H.; Neumann, S.; Beller, M. Chem.—Eur. J. 2012, 18, 13619.
- (11) For examples of transition-metal catalyzed C—H bond arylation, see: (a) Ackermann, L.; Barfusser, S.; Kornhaass, C.; Kapdi, A. R. *Org. Lett.* **2011**, *13*, 3082. (b) Lewis, J. C.; Berman, A. M.; Bergman, R. G.; Ellman, J. A. *J. Am. Chem. Soc.* **2008**, *130*, 2493. (c) Lewis, J. C.; Wiedemann, S. H.; Bergman, R. G.; Ellman, J. A. *Org. Lett.* **2004**, *6*, 35.
- (12) (a) Mei, L.; Hai, Z. J.; Jie, S.; Ming, Z. S.; Hao, Y.; Liang, H. K. J. Comb. Chem. 2009, 11, 220. (b) Mohammadpoor-Baltork, I.; Khosropour, A. R.; Hojati, S. F. Synlett 2005, 18, 2747. (c) Mohammadpoor-Baltork, I.; Khosropour, A. R.; Hojati, S. F. Catal. Commun. 2007, 8, 200. (d) Jnaneshwara, G. K.; Deshpande, V. H.; Lalithambika, M.; Ravindranathan, T.; Bedekar, A. V. Tetrahedron Lett. 1998, 39, 459. (e) Shaabani, A.; Seyyedhamzeh, M.; Maleki, A.; Rezazadeh, F. Appl. Catal., A 2009, 358, 146. (f) Mohammadpoor-Baltork, I.; Mirkhani, V.; Moghadam, M.; Tangestaninejad, S.; Zolfigol, M. A.; Abdollahi-Alibeik, M.; Khosropour, A. R.; Kargar, H.; Hojati, S. F. Catal. Commun. 2008, 9, 894. (g) Mohammadpoor-Baltork, I.; Moghadam, M.; Tangestaninejad, S.; Mirkhani, V.; Hojati, S. F. Catal. Commun. 2008, 9, 1153. (h) Wang, L.; Guo, B.; Li, H.-X.; Li, Q.; Lia, H.-Y.; Lang, J.-P. Dalton Trans. 2013, 42, 15570. (i) Li, X. N.; Zhou, B. Y.; Zhang, J.; She, M. Y.; An, S. J.; Ge, H. X.; Li, C.; Yin, B.; Li, J. L.; Shi, Z. Eur. J. Org. Chem. 2012, 8, 1626. (j) Ge, H. X.; Liu, P.; Li, X. N.; Sun, W.; Li, J. L.; Yang, B. Q.; Shi, Z. Tetrahedron 2013, 69, 6591.
- (13) Garg, P.; Milton, M. D. Tetrahedron Lett. 2013, 54, 7074.
- (14) (a) Wuest, W. M.; Sattely, E. S.; Walsh, C. T. *J. Am. Chem. Soc.* **2009**, *131*, 5056. (b) Castellano, S.; Kuck, D.; Sala, M.; Novellino, E.; Lyko, F.; Sbardella, G. *J. Med. Chem.* **2008**, *51*, 2321. (c) Munoz, B.;

- Stearns, B.; Vernier, J. M.; Wang, B.; Bonnefous, C.; Zhao, X.; Arruda, J.; Campbell, B. T.; Cube, R. V. Patent no. US 20050065340, 2005.
- (15) (a) Lester, R. P.; Camp, J. E. ACS Sustainable Chem. Eng. 2013, 1, 545. (b) Voss, M. E.; Beer, C. M.; Mitchell, S. A.; Blomgren, P. A.; Zhichkin, P. E. Tetrahedron 2008, 64, 645. (c) Schaefer, F. C.; Peters, G. A. J. Org. Chem. 1961, 26, 412. (d) Roger, R.; Neilson, D. G. Chem. Rev. 1961, 61, 179. (e) Mahadevan, S.; Venkatasubban, K. Patent no. WO 2012148624 A1 20121101, 2012.
- (16) (a) Patel, M. B.; Modi, N. R.; Raval, J. P.; Menon, S. K. Org. Biomol. Chem. **2010**, 10, 1785. (b) Blois, M. S. Nature **1958**, 181, 1199.
- (17) (a) MacLean, P. D.; Chapman, E. E.; Dobrowolski, S. L.; Thompson, A.; Barclay, L. R. C. *J. Org. Chem.* **2008**, 73, 6623. (b) Wang, D.; Hu, X.-S.; Zhao, G. H. *Int. J. Food Sci. Technol.* **2008**, 43, 1880.
- (18) Leung, S. C.; Gibbons, P.; Amewu, R.; Nixon, G. L.; Pidathala, C.; Hong, W. D.; Pacorel, B.; Berry, N. G.; Sharma, R.; Stocks, P. A.; Srivastava, A.; Shone, A. E.; Charoensutthivarakul, S.; Taylor, L.; Berger, O.; Mbekeani, A.; Hill, A.; Fisher, N. E.; Warman, A. J.; Biagini, G. A.; Ward, S. A.; O'Neill, P. M. *J. Med. Chem.* **2012**, *55*, 1844.
- (19) Molander, G. A.; Gormisky, P. E.; Sandrock, D. L. J. Org. Chem. **2008**, 73, 2052.
- (20) Gemma, S.; Camodeca, C.; Brindisi, M.; Brogi, S.; Kukreja, G.; Kunjir, S.; Gabellieri, E.; Lucantoni, L.; Habluetzel, A.; Taramelli, D.; Basilico, N.; Gualdani, R.; Tadini-Buoninsegni, F.; Bartolommei, G.; Moncelli, M. R.; Martin, R. E.; Summers, R. L.; Lamponi, S.; Savini, L.; Fiorini, I.; Valoti, M.; Novellino, E.; Campiani, G.; Butini, S. *J. Med. Chem.* 2012, 55, 10387.
- (21) Molander, G. A.; Ryu, D.; Hosseini-Sarvari, M.; Devulapally, R.; Seapy, D. G. J. Org. Chem. 2013, 78, 6648.
- (22) Dinsmore, C. J.; Zartman, C. B.; Baginsky, W. F.; O'Neill, T. J.; Koblan, K. S.; Chen, I.-W.; McLoughlin, D. A.; Olah, T. V.; Huff, J. R. Org. Lett. 2000, 2, 3473.
- (23) Aakeroy, C. B.; Desper, J.; Urbina, J. F. Cryst. Growth Des. 2005, 5, 1283.
- (24) Keil, S.; Urmann, M.; Glien, M.; Wendler, W.; Chandross, K.; Lee, L. Patent no. WO2007039176, 2007.
- (25) Rizzo, S.; Bartolini, M.; Ceccarini, L.; Piazzi, L.; Gobbi, S.; Cavalli, A.; Recanatini, M.; Andrisano, V.; Rampa, A. *Bioorg. Med. Chem.* **2010**, *18*, 1749.
- (26) Tokizane, M.; Sato, K.; Sakami, Y.; Imori, Y.; Matsuo, C.; Ohta, T.; Ito, Y. Synthesis 2010, 36.
- (27) Guo, D.-C.; Li, P.-L.; Wang, X.; Wang, L.-Y.; Wu, P.-L. Synth. Commun. 2010, 40, 3315.
- (28) Gaster, L. M.; King, F. D.; Wyman, P. A. Patent no. USS972951A, 1999.
- (29) Tsuda, Y.; Kuriyama, M.; Onomura, O. *Chem.—Eur. J.* **2012**, *18*, 2481.
- (30) Croisy-Delcey, M.; Bisagni, E. J. Chem. Soc., Chem. Commun. 1984, 897.
- (31) Barnard, J. H.; Wang, C.; Berry, N. G.; Xiao, J.-L. Chem. Sci. 2013, 4, 1234.
- (32) Sahnoun, S.; Messaoudi, S.; Peyrat, J.-F.; Brion, J.-D.; Alami, M. Tetrahedron Lett. 2012, 53, 2860.
- (33) Kita, Y.; Nishii, Y.; Higuchi, T.; Mashima, K. Angew. Chem. Int. Ed. 2012, 51, 5723.